



ZEISS Primo Star iLED

Selected Fluorescence Applications in Laboratories
and Education



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Primo Star iLED is your flexible solution with LED fluorescence excitation and transmitted light brightfield illumination. Analyze tuberculosis with Ziehl-Neelsen staining in brightfield or use fluorescence excitation with Auramine O dye. In combination with new iLED fluorescence attachments: even greater versatility can now be achieved with Primo Star iLED for many fluorescent labels. Primo Star iLED allows you to switch easily between the two modes.

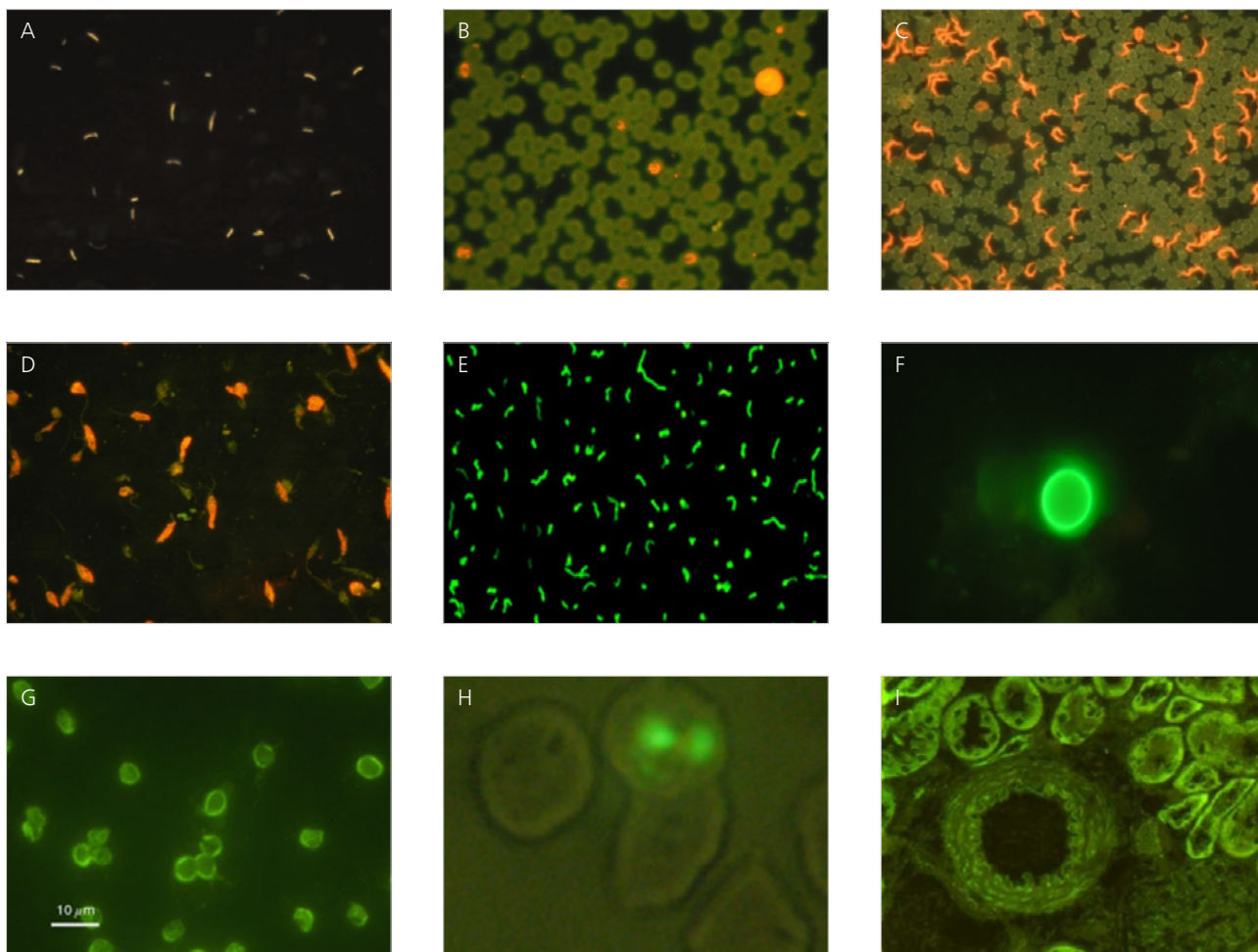
Applications**	Brightfield (BF) ***	LED fluorescence (FL)		Examples [detection of]
		Dye	Filter set (FS) & LED	
Tuberculosis diagnosis	Ziehl-Neelsen stain	Auramine O*	FS 67, LED 455 nm	Mycobacteria (BF, FL ¹⁻¹⁰)
Malaria diagnosis	Giemsa's stain	Acridine orange*	FS 67, LED 455 nm	Protozoa (BF, FL ¹¹⁻¹³)
Sleeping Sickness diagnosis	Giemsa's stain	Acridine orange	FS 67, LED 455 nm	<i>Trypanosoma brucei</i> (BF, FL ¹³⁻¹⁵)
Leishmaniasis diagnosis	Leishman's or Giemsa's stain	Acridine orange	FS 67, LED 455 nm	Leishmania parasites (BF, FL ¹³)
With optional accessories (Filter sets & LEDs):				
Gastritis diagnosis	H & E stain	FITC	FS 09, LED 470 nm	Tissue morphology (BF), <i>Helicobacter pylori</i> (FL)
Giardiasis diagnosis		FITC or SYBR-Green / SYBR-Gold	FS 09, LED 470 nm	<i>Giardia lamblia</i> (FL) ¹⁶
Cryptosporidiosis diagnosis	H & E stain	FITC or SYBR-Green / SYBR-Gold	FS 09, LED 470 nm	Tissue morphology (BF), Cryptosporidia (FL) ¹⁷
Malaria diagnosis	Giemsa's stain	SYBR-Green	FS 09, LED 470 nm	Protozoa (BF, FL)
Histology / Immunohistology	H & E stain	FITC	FS 09, LED 470 nm	Tissue morphology (BF, FL)

* No coverslip needed, observed at 400 X magnification.

** For the best use of Primo Star iLED that comes with FS 67 and LED 455: equipped with a 40x objective D=0 and a 100x objective D=0.17.

*** Please take note that the references are for fluorescence application only (iLED part).

Application examples:

**Short notes:**

- A. *Tubercle bacilli*
- B. *Plasmodium knowlesi*
- C. *Trypanosomes*
- D. *Leishmania donovani* promastigotes
- E. *Helicobacter pylori*
- F. *Giardia*
- G. *Cryptosporidia*
- H. *Plasmodium falciparum*
- I. *Mouse capillary*

Legend:

(A) *Tubercle bacilli* stained with auramine O (courtesy of CDC). (B) Malaria parasites (small orange structures) inside red blood cells (green) of a baboon experimentally infected with *Plasmodium knowlesi* and stained with acridine orange. White blood cells also stain orange (blood smear courtesy of Dr. Maina Ngotho, Institute of Primate Research, Nairobi). (C) A thin blood smear stained with acridine orange showing trypanosomes (orange) alongside red blood cells (green). (D) Cultured *Leishmania donovani* promastigotes (orange with flagella) stained with acridine orange (slide courtesy of Dr. Maina Ngotho).¹³ (E) *Helicobacter pylori* immuno-labeled with FITC. (F) *Giardia* from contaminated water immuno-labeled with FITC (courtesy of Dr. H.P. Fuechslin, Bachema AG). (G) *Cryptosporidia* from contaminated drinking water immuno-labeled with FITC (courtesy of Mr. Brian Oram, Wilkes University).¹⁷ (H) *Plasmodium falciparum* stained with SYBR Green I (courtesy of Dr. West Suhanic¹¹). (I) Mouse capillary immuno-labeled with FITC.

Highlights

Use ZEISS Primo Star in combination with iLED fluorescence attachments for many fluorescent labels and profit from.

- fast and efficient testing
- reflected-light fluorescence (FL) together with transmitted-light brightfield (BF)
- easy switching between FL excitation and BF illumination
- economical LED concept: long-lasting, retrofittable with any Primo Star
- versatility with options for fluorescence attachments
- battery pack for operation without a main power supply
- special eyecups eliminate the need for a dark room during a test for tuberculosis, malaria, sleeping sickness, or leishmaniasis
- easy to operate
- durable and robust
- tried-and-tested ZEISS optics made from high-quality glass
- high-quality materials
- worldwide support from ZEISS

References:

- [1] Turnbull, E.R. et al. An evaluation of the performance and acceptability of three LED fluorescent microscopes in Zambia: lessons learnt for scale-up. *PLoS One* 6, e27125 (2011).
- [2] Minion, J., Pai, M., Ramsay, A., Menzies, D. & Greenaway, C. Comparison of LED and Conventional Fluorescence Microscopy for Detection of Acid Fast Bacilli in a Low-Incidence Setting. *PLoS ONE* 6, e22495 (2011).
- [3] Drobniowski, F., Nikolayevskyy, V., Balabanova, Y., Bang, D. & Papaventsis, D. Diagnosis of tuberculosis and drug resistance: what can new tools bring us? *The international journal of tuberculosis and lung disease: the official journal of the International Union against Tuberculosis and Lung Disease* 16, 860-870 (2012).
- [4] Albert, H. et al. Feasibility of magnetic bead technology for concentration of mycobacteria in sputum prior to fluorescence microscopy. *BMC infectious diseases* 11, 125 (2011).
- [5] Parsons, L.M. et al. Laboratory diagnosis of tuberculosis in resource-poor countries: challenges and opportunities. *Clinical microbiology reviews* 24, 314-350 (2011).
- [6] Minion, J., Sohn, H. & Pai, M. Light-emitting diode technologies for TB diagnosis: what is on the market? *Expert Review of Medical Devices* 6, 341-345 (2009).
- [7] Alfred, N. et al. Optimising Mycobacterium tuberculosis detection in resource limited settings. *BMJ open* 4, e004093 (2014).
- [8] Albert, H. et al. Performance of three LED-based fluorescence microscopy systems for detection of tuberculosis in Uganda. *PLoS One* 5, e15206 (2010).
- [9] Henostroza, G. et al. The high burden of tuberculosis (TB) and human immunodeficiency virus (HIV) in a large Zambian prison: a public health alert. *PLoS One* 8, e67338 (2013).
- [10] Ramsay, A., Steingart, K.R., Cunningham, J. & Pai, M. Translating tuberculosis research into global policies: the example of an international collaboration on diagnostics. *The international journal of tuberculosis and lung disease : the official journal of the International Union against Tuberculosis and Lung Disease* 15, 1283-1293 (2011).
- [11] Suhanic, W., Crandall, I. & Pennefather, P. An informatics model for guiding assembly of telemicrobiology workstations for malaria collaborative diagnostics using commodity products and open-source software. *Malaria journal* 8, 164 (2009).
- [12] Lenz, D. et al. Assessment of LED fluorescence microscopy for the diagnosis of Plasmodium falciparum infections in Gabon. *Malaria journal* 10, 194 (2011).
- [13] Ndung'u, J.M., Bieler, S. & Roscigno, G. "Piggy-backing" on diagnostic platforms brings hope to neglected diseases: the case of sleeping sickness. *PLoS neglected tropical diseases* 4, e715 (2010).
- [14] Diagnostics, F.f.I.N. Practical fluorescence microscopy for detection of trypanosomes. (2010).
- [15] Matovu, E., Kazibwe, A.J., Mugasa, C.M., Ndungu, J.M. & Njiru, Z.K. Towards Point-of-Care Diagnostic and Staging Tools for Human African Trypanosomiasis. *Journal of tropical medicine* 2012, 340538 (2012).
- [16] Keserue, H.A. et al. Comparison of rapid methods for detection of Giardia spp. and Cryptosporidium spp. (oo)cysts using transportable instrumentation in a field deployment. *Environmental science & technology* 46, 8952-8959 (2012).
- [17] <http://www.water-research.net/cryptosporidium.htm>
- [18] http://www.mycology.adelaide.edu.au/Laboratory_Methods/Microscopy_Techniques_and_Stains/calcofluor.html



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